

# Development of Native and Denaturing LC-MS Methods for Characterization of Cysteine-Linked Antibody Drug Conjugates Using a Non-Toxic Mimic

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## Introduction

Here we utilize a cysteine-linked antibody fluorophore conjugate to show how LC-MS methods may be optimized using a non-toxic ADC-mimic. The ADC-mimic was used to develop native and reduced SEC-MS conditions. The reduced mimic's heavy and light chains, as well as its IdeS-proteolysis fragments, were used to develop reversed-phase LC-MS methods. A tryptic digest of the ADC mimic was used to develop a peptide mapping LC-MS method.

## Methods

### Native SEC-MS

Column: Tosoh TSK Gel SW3000XL, 2.0 mm x 300 mm, 4 μm; 25 °C  
Isocratic flow: 100 mM ammonium acetate, pH 7, 70 μL/min  
Handling: PNGase F 37 °C, 18 h

### Reduced SEC-MS

Column: Tosoh TSK Gel SW3000XL, 2.0 mm x 300 mm, 4 μm; 25 °C  
Isocratic flow: 30% ACN 0.1% TFA, 70 μL/min  
Handling: PNGase F 37 °C 18 h; 50 mM DTT, 37 °C, 1 h

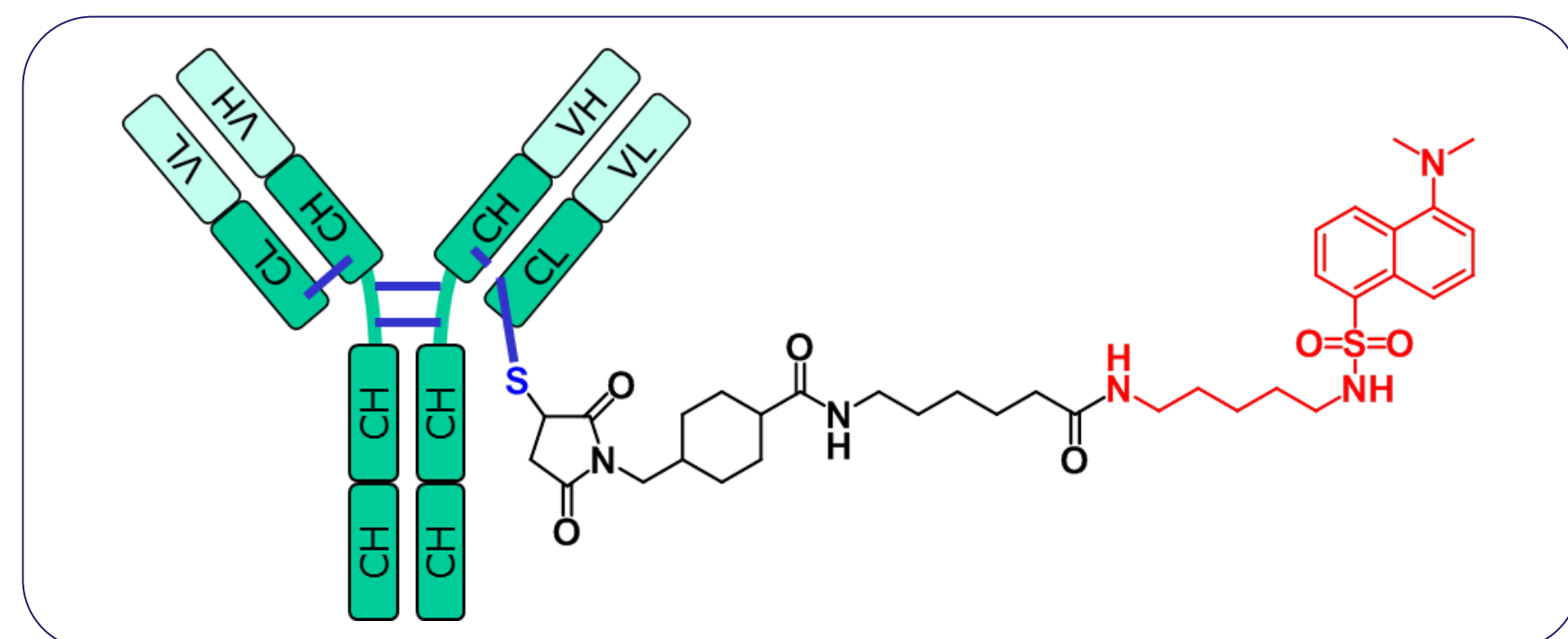
### Antibody Subunit RPLC-MS

Column: BIOshell A400 Protein C4, 300 μm x 150 mm, 3.4 μm; 65 °C  
Phases: Water, 0.1% TFA; ACN, 0.1% TFA  
Gradient: Desalt 5% organic; 31%-32.5% organic / 4 min; 35%-38% organic / 10 min, 40 μL/min  
Handling: 7.4 M GuHCl / 100 mM DTT 50 °C 1 h; optionally IdeS 37 °C 3 h

### Peptide Mapping

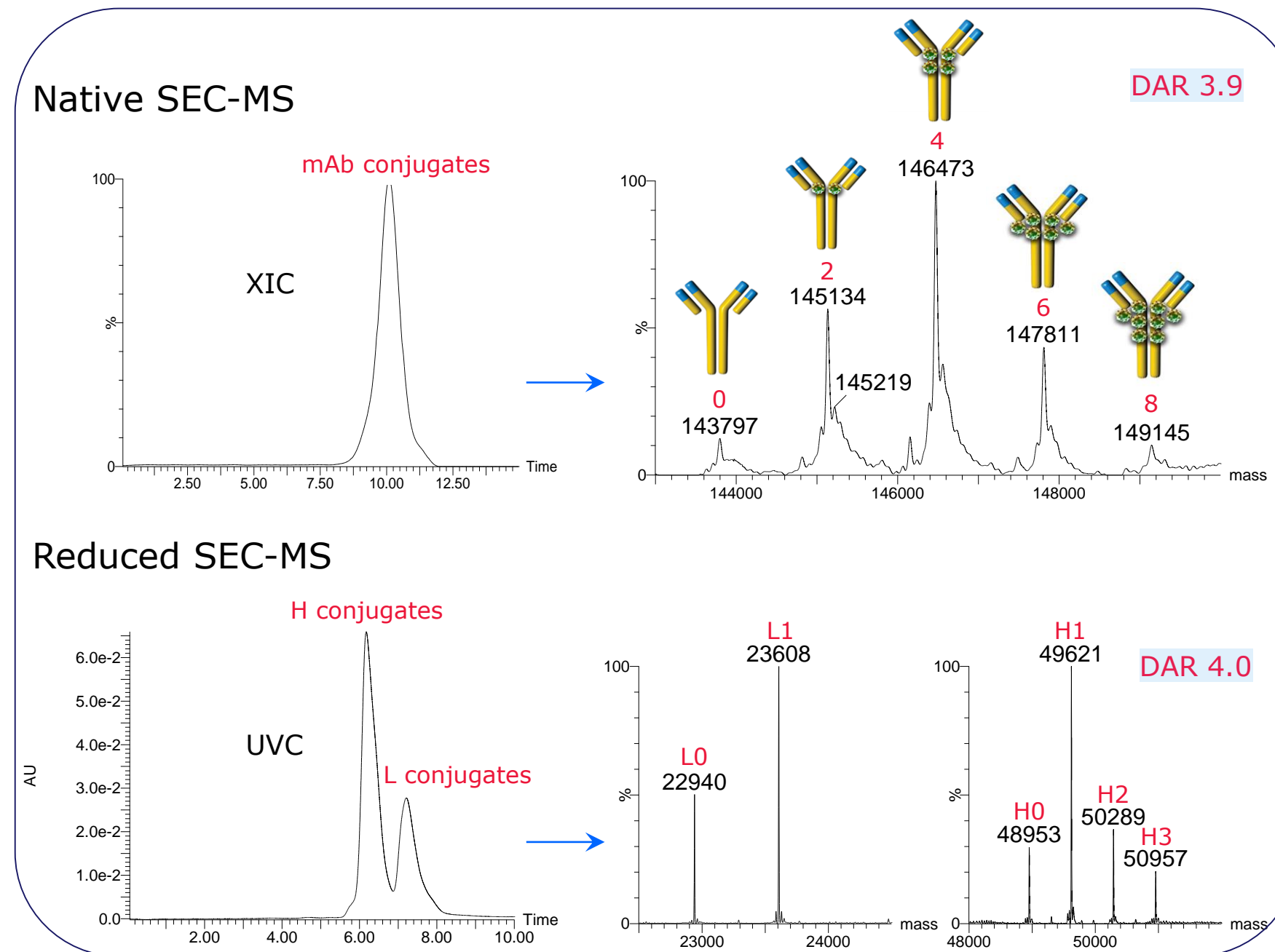
Column: BIOshell A160 C18, 300 μm x 300 mm, 2.7 μm; 40 °C  
Phases: Water, 0.1% FA; ACN, 0.1% FA  
Gradient: 3%-57% organic / 120 min; 10 μL/min  
Handling: Urea/DOC; TCEP/IAA; FASP digestion with Trypsin/LysC, 37 °C 16 h

An M-Class Acquity-Xevo G2 QToF (Waters) was used in all methods.

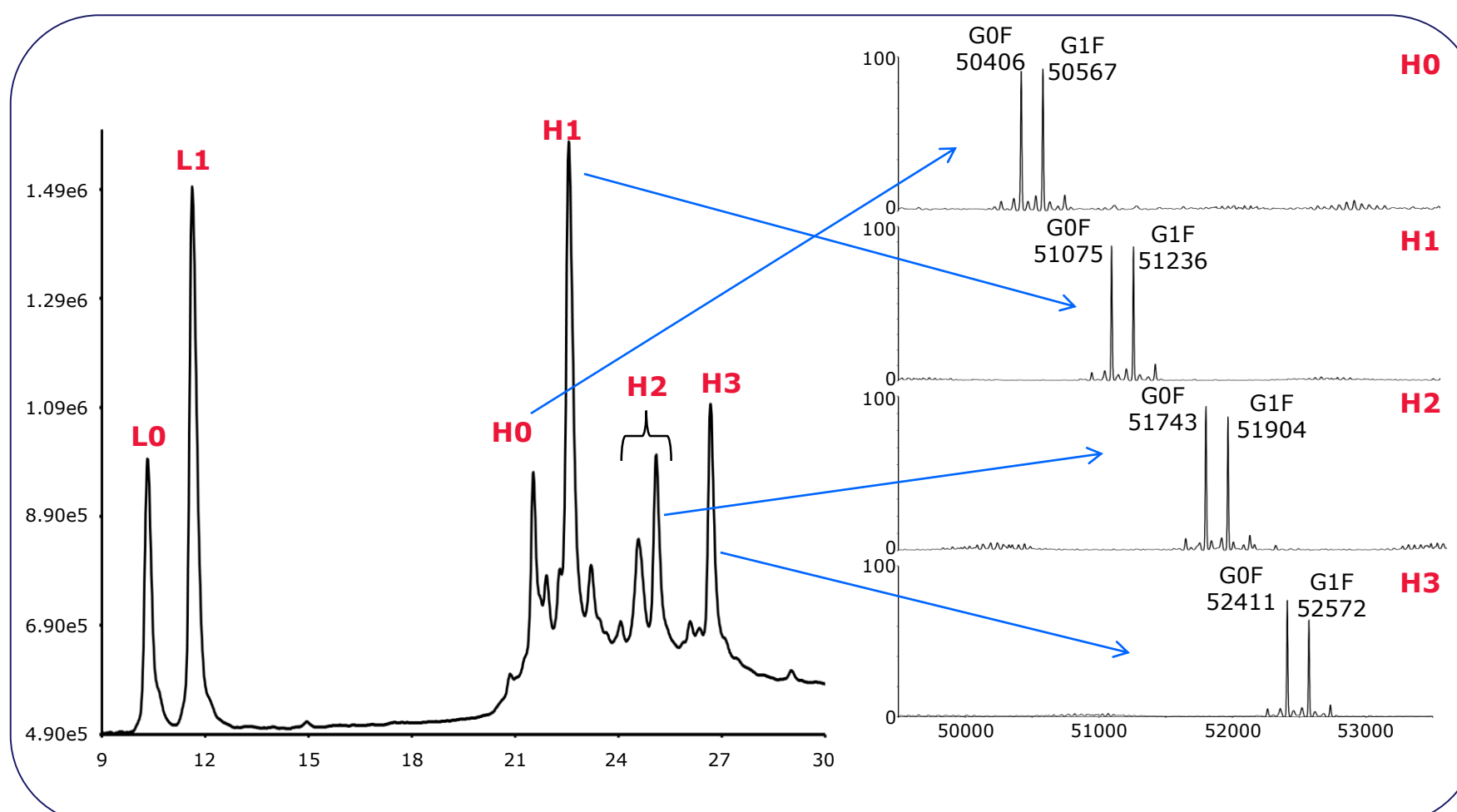


**Figure 1.** Structure of the SigmaMAB Antibody Drug Conjugate (ADC) Mimic, a human IgG1 monoclonal antibody with 0-8 dansyl fluorophores conjugated via an LC-SMCC crosslinker. Each dansyl-LC-SMCC attachment imparts a mass shift of 668 Da.

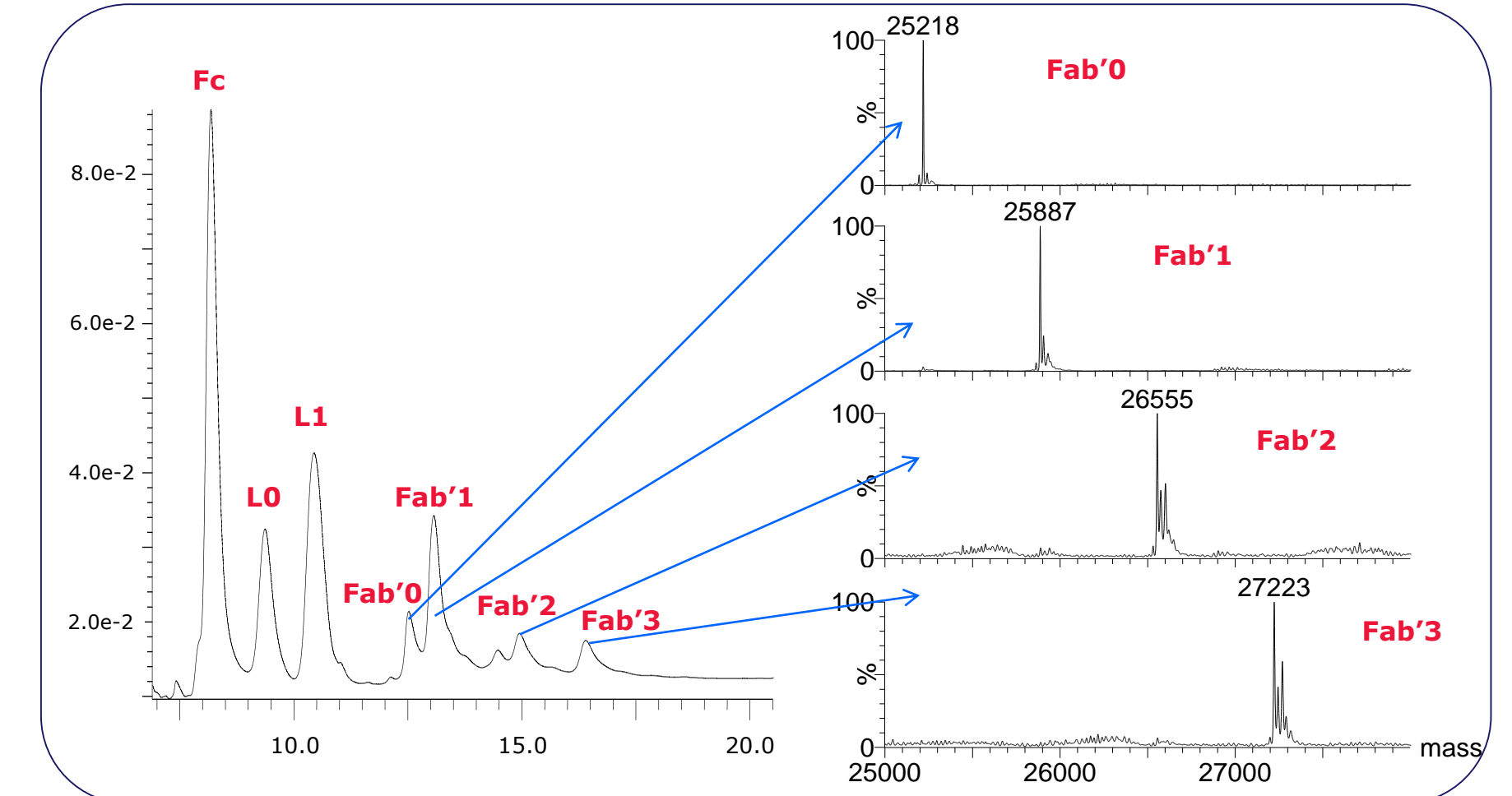
## Results



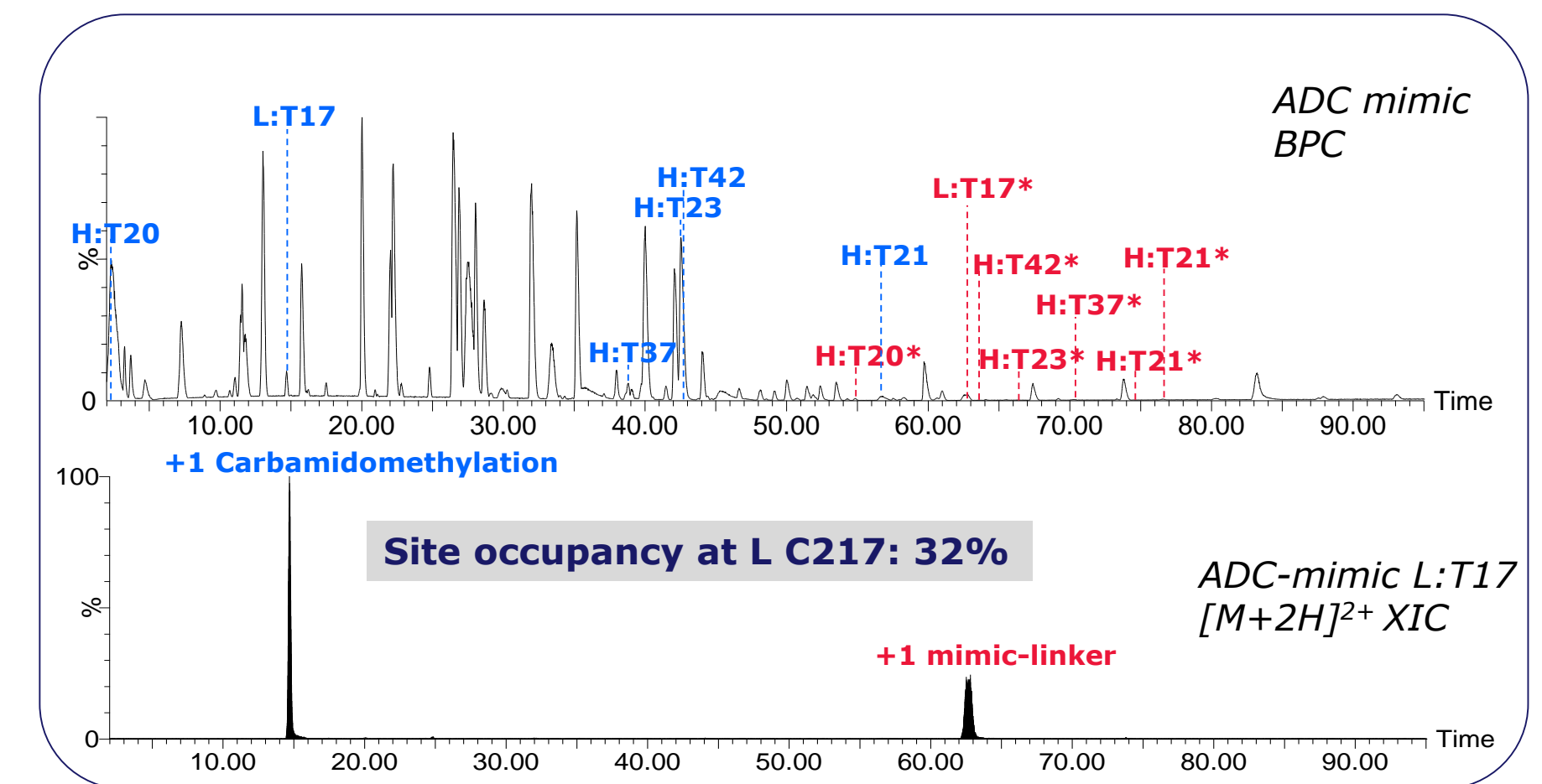
**Figure 2.** Deglycosylated ADC-mimic characterization by native and reduced SEC-MS methods. Each annotation counts the number of mimic-linkers conjugated to the antibody or subunit.



**Figure 3.** RPLC total ion chromatogram and deconvoluted MS spectra of reduced ADC-mimic heavy chain species. Annotation indicates glycoform and number of mimic-linker attachments.



**Figure 4.** RPLC UV chromatogram and deconvoluted MS spectra of IdeS/reduced ADC-mimic Fab' species. Each annotation counts the number of mimic-linkers conjugated to the antibody subunit.



**Figure 5.** Tryptic peptide map of the ADC-mimic. Red and blue annotations are of conjugated and unconjugated peptides, respectively. Site occupancy was determined from XIC peak area ratio.

## Conclusions

We have utilized a non-toxic ADC-mimic, with Cys-conjugation to dansyl-cadaverine-SMCC, to develop native and reduced SEC-MS methods, subunit RPLC-MS methods, and a peptide mapping method.